

European Journal of Nutrition & Food Safety

Volume 16, Issue 1, Page 66-78, 2024; Article no.EJNFS.111129 ISSN: 2347-5641

Ameliorative Activities of Morus and Jamun against Cr Induced Andro-Hepatic Anomalies

Tahir Abbas ^{a*}, Khawaja Raees Ahmad ^{b++}, Asmatullah ^{c#}, Haider Ali Akhtar ^d and Tehreem Fatima ^e

^a Principal Government College, Kotmomin, Sargodha, Pakistan. ^b Department of Zoology, Sargodha University, Sargodha, Pakistan. ^c Department of Zoology, University of The Punjab, Lahore Pakistan. ^d Department of Zoology, University of Central Punjab, Sargodha Campus, Pakistan. ^e Isfahan University of Medical Science, Iran.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/EJNFS/2024/v16i11379

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/111129

> Received: 05/11/2023 Accepted: 09/01/2024 Published: 24/01/2024

Original Research Article

ABSTRACT

Chromium (Cr) is a vital micronutrient for sugar metabolism while its unauthorized use may agonize health fatalities. This study was conducted about the novelty about exposure of hexavalent chromium (Cr⁺⁶) histopatholgical potentials along with Mulberry Fruit Extract and Jamun Fruit Extract to define the ameliorative aptitude. The histological finding in Cr exposure specify; cessation of testicular seminiferous tubules (ST), annihilation of smooth muscles in basement membrane, Leydig's cell and spermatids. The ST have halted spermatogenesis, with irregular boundary and dead spermatogonia and Leydig's cells are disorganized. The sperm head

⁺⁺ Professor;

[#] Associate Professor;

^{*}Corresponding author: Email: tahirabbasbloch@gmail.com;

Eur. J. Nutr. Food. Saf., vol. 16, no. 1, pp. 66-78, 2024

cross sectional area (CSA), tail length and middle piece diameter ($p \le 0.001$) significantly reduced while there is significant elevation of hepatocytes nuclei size, central vein CSA and mean width of Sinusoidal Spaces as compared to control. The protuberant marks of steatosis, fibrosis, dehydration and atrophy were nullified by designated fruit extracts and their possible bio-chelating effects was obvious due to the presence of anthocyanin, β -sitosterol and phytochemicals. The given results specify that hexavalent Cr induce andro-hepatic anomalies when it is freely used without scientific authentication. Cr⁺⁶ as food additives in coloring rice and local sweets should be prohibited while bio-products of Morus and Jamun can be recommended and sponsored for traditional medicines.

Keywords: Chromium; steatosis; fibrosis; dehydration and atrophy.

LIST OF ABBREVIATIONS

Abbreviations	Abbreviations details
ALP	alkaline phosphatase
ALT	alanine aminotransferase
AMPK	amp-activated protein kinase
AS	attached sperm
AST	aspartate aminotransferase
BUN	Blood Urea Nitrogen
СН	club headed sperm
CSA	cross sectional area
C.V	central vein
Cr	Chromium
Cr-M	chromium+ Mulberry
Cr-J	chromium+Jambul
Cr-MJ	chromium+ Mulberry +Jambul
CRT	Creatine
DS	dislocated sperm
FSH	follicle stimulating hormone
GLU	glucose (sugar)
Hb	Hemoglobin
HDL	high density lipoprotein
He.C	Hepatocytes
H.N	hepatocytic nucleus
JFE	Jambul Fruit Extract
K.C	kupffer cells
LDL	low density lipoprotein
MCV	mean cell volume
MFE	Mulberry Fruit Extract
P. SPER	primary spermatocytes
PBH	parrot beak headed sperm
RBC	erythrocyte count
ROS	reactive oxygen species
S. GONIA	Spermatogonia
SGOT	serum glutamic oxalacetic transaminase
SGPT	serum glutamic pyruvate transaminase
S.S	sinusoidal spaces
ST	seminiferous tubules
ТА	total albumin
TG	Triglyceride
TP	total protein
VLDL	very low density lipoprotein
UA	Uric Acid
WBC	white blood cell count

1. INTRODUCTION

The unhygienic heavy metals contaminated water frequently used to irrigate crops and bioproducts of such crops cause inevitable anomalies [1]. Metals such as lead, mercury, chromium, arsenic, copper, cadmium, and iron in contaminated water and food supplements generate reactive oxygen species [2,3], affect immune system and body organs such as testes, kidneys and liver [4]. The adequate amount of chromium (Cr) as food supplement enhance endocrine system and ethological activities [5] but their ridiculous use may cause severe injuries on androgen receptors lead hepatic, renal and hematologic anomalies [6]. The elevated androgen block endogenous androgens of Leydig cells [7] and effect testicular seminiferous tubules and hepatocytes which fight against reactive oxygen species (ROS) in liver [8,9].

Many herbs are investigated as source of antioxidants as free radical's scavenger [27] and ameliorative competency against heavy metals [28,29,30] Antioxidant may be synthesized in the body or obtained in the diet; have capability of βcarotene-linoleate and β-sitosterol with reducing superoxide, nitric oxide-scavenging power, capacity and ferrous ion chelating potency [31] and their anthocyanin control lipid peroxidation [32]. The flavonoids and polyphenols in

Trianthema triquetra [33], *Pisonia aculeata* [34], *Benincasa hispida* and Castor oil [35] are very beneficial in scavenging the free radicals [36] by regulating aspartate aminotransferase, alanine aminotransferase, total serum bilirubin and malondialdehyde in hepatocytes [37].

Mulberry antioxidants with cytotoxic protective activitv reduce edema [38], also have ameliorative capability against hyperlipidemia, lipogenesis and fatty acid oxidation [39]. Morus nigra have anti-cancerous, anti-inflammatory, antibacterial and anti-fungal, antimicrobial and radical rummaging activities [40]. Their quercetin from flavonoids protects lipid peroxidation [41]. Pharmacologically Syzigium cumini fruit extracts protect the cultured human peripheral blood lymphocytes from DNA damage [42] also recommended to treat human breast repair hepatocyte from hydrogen cancer. peroxide injuries and gamma-irradiations [43]. Their anthocyanins are anti-spasmodic [44] can ameliorate hepatic enzymes to moderate serum ALT and AST against methylmercury [45]. The flavonoids in litchi (Litchi chinensis) are effective against cancer [46], Viola odorata for asthma [47], red clover for hot flashes/menopausal symptoms [48], green tea for the risk of prostate problems [49], cactus (Opuntia ficus-indica) cladodes on methotrexate-induced oxidative

Plant	Compound	Bio-chelation	References
Fruit extract	Anthocyanin	Fe	[10,11]
Mulberry extracts	phenolics, β-	CCl ₄	[12,13]
	sitosterol	and heavy	
	and	metals	
	anthocyanins		
Morus alba	Phytochemicals	CCl ₄	[14]
Syzygium cumini	Antioxidant,	Nitric oxide, Fe,	[15,16]
	anthocyanin	CCl ₄	
	Flavonoid	Fe,	
		methylmercury	
Moringa oleifera	Phytochemicals	Cr	[17,18]
Hippophae rhamnoides			
Plants, fruits, vegetable	Vitamins E/C	Heavy metals	[19,20]
		Pb	
Mangifera indica	Phytochemicals	Fluoride	[21,22]
Red cabbage		Heavy metals	
Plants	L-carnitine	CCl4	[23,24]
Salvia plebeia	Flavonoid		
Plants	Ascorbic acid,	Arsenic	[25]
	tocopherol		
Plants derived drug	Deferiprone	Fe, Al	[26]

Table. 1. Plants phytochemical compounds used for bio-chelation

damage [50] and chamomile (*Matricaria recutita*) as antioxidant and anticancer activities [51]. The enzymatic changes and gene expressions are almost same in humans and rodents [52] so mice mammalian model was selected for this study, and the cheapest economical fruit Morus and Jamun were selected to probe the poisonous effects of heavy metals and shield against androhepatic anomalies at micrometric level.

2. MATERIALS AND METHODS

The study was conducted on 25-30g/3-4months old 50 male albino laboratory mice (*Mus musculus*), kept at 26 ± 4 C°/45% relative humidity with 12-hr L/D cycles throughout experimental duration.

2.1 Preparation of Solution and Fruit Extracts

Dose for various groups were prepared from pure Potassium dichromate ($K_2 Cr_2 O_7$), stock solution of 1000ppm and diluted up-to 50ppm required solutions. Ripe black berry of *Morus nigra* and *Syzygium cumini* were washed, and 100g of their pulp was blended in an electric juicer in 100ml cooled boiled drinking water for 5minutes and finally centrifuged at 500rpm for 10minutes. The supernatant was immediately placed in sterilized 5ml capacity ice-cube dishes, store in sterilized plastic bags at -30°C and a fresh thawed cube was used for each experiment at room temperature.

2.2 Experimental Groups

Animals were randomly divided into 5 groups (n=10) as: Control group (C); provided boiled

cooled mineral water for 15days, Cr-group (Cr); provided 50ppm Cr-solution for first 10days at *ad libitum* followed by simple mineral water for next 5days, Cr +Mulberry group (Cr-M); as Cr group but the last 5days they were given MFE, similarly Cr +Jamun group (Cr-J) given JFE and Cr +Mulberry+Jamun group (Cr-MJ) were given equally mixed MFE and JFE through gavage as post-treatment 0.25ml/12hrs for next 5 days.

2.3 Histological and Micrometric Analysis

The animals were euthanized by cervical dislocation and dissected out on day 16th for histological hematoxylin and eosin (HE) staining, testicular smear formation and micrometric analysis following [74] protocol. Photograph at 100x, 400x and 1000x with 7.2 MP digital camera (Sony) were processed in CoreIDRAW11 for micrometry. The results were expressed as mean \pm SD and the data was analyzed through ANOVA (two factors, without replication) and Duncan's Multiple Range Test (post hoc analysis).

3. RESULTS

In control group testes of animals specify the clear boundaries of seminiferous tubules (ST) with integral basement membranes, spermatogonia (Sp.g) with multiple whirls of spermatocytes (Sp.c) and secondary spermatocytes (Se.c). The sperms with parrot beak headed (PBH) and club headed (CH) along with prominent elongated tail (Sp.T) were clearly visible. All whirls are equally and systematically are arranged and there was no sign of anomalies in all animals of control group (Fig. 1).



Fig. 1. Mice Seminiferous Tubule, control group Sp.g; spermatogonium, Se.c; sertoli cell, Sp.c; spermatocyte, PBH; parrot beak headed spermatozoa, CH; club headed spermatozoa, Sp.T; Spermatozoa Tail

The seminiferous tubules (ST) in animals without Cr treatment were symmetrical equally distributed rounded with prominent Leydig's cells (Fig. 2-a), spermeogenesis (Fig. 2-b) and spermatozoa with elongated tails directed towards lumen the (Fig. 2-c). The spermatogonium with prominent dark stain nuclei without vacuolation and cracks (Fig. 2-d). In Cr treated group there was breakdown of basement membrane, destruction of smooth muscles, Levdig's cell (Fig. 2-e) and spermatid without spermeogenesis (Fig. 2-f). The tailless sperm (Fig. 2-g) and debris filled in ST lumen appear like vortex (Fig. 2-h). The ST with halted spermatogenesis, with irregular boundary and basement membrane loses inter-tubular junction (Fig. 2-i). The dead spermatogonia and Leydig's cells were disorganized (Fig. 2-j, I) and whirls of primary and secondary spermatocytes lost their regular symmetry (Fig. 2-k).

Morus as post-treatment ameliorate the basement membrane of ST as shown in Fig. 2-CrM-m. There are prominent Sertoli cells (Fig. 2-CrM-n) after treatment indicate clear systematic spermatogenesis (Fig. 2-CrM-o). The regular whirls of Sp.g, Se.c, and Sp.c, with healthy PBH, CH and Sp.T (Fig. 2-CrM-p in Morus posttreatment specify prominent recovery of testicular anomalies.

Similarly Jamun improve the Leydig's cell, basement membrane along with smooth muscles around vacuolated regions (Fig. 2-CrJ-r). There is reorganization of spermeogenesis (Fig 2-CrJs) and improvement in primary and secondary spermatocytes whirls (Fig. 2-CrJ-t,u) after Jamun treatment. The Morus and Jamun co-treatment synchronized their effects by reducing vacuolated cells and removing debris. The reorganizing cellular texture and symmetrical whirls of ST (Fig. 2-CrJM-w,x,y,z) also intimate the ameliorative aptitude.

The testicular and seminal vesicle smear of normal animals as control group with PBH along with long tail; without any kink (Fig. 3-1, 5). In Cr treatment sperm show kink in their tails and weak connection b/w sperm head and tail (Fig. 3-2,7); sperm heads are frequently distributed specify halted spermatogesis and spermeogenesis (Figs. 3-8,9), while slides (Figs.3-3,4,6) indicate Morus and Jamun amelioration.

3.1 Micrometric Histological Study of Mice Testes

3.1.1 Sperm micrometric study

The mean CSA of head, length of tail and diameter of mid piece was measured from testicular smear as;

3.1.2 Micrometric histological study of mice liver

The cross sectional of liver indicate destruction of hepatocytes in Cr treated group (Fig. 4). There is significant ($p \le 0.001$) elevation of hepatocytes nuclei size, central vein CSA and mean width (μ m) of Sinusoidal Spaces (S.S) after Cr exposure. The mean number of hepatocytes and relative area occupied by hepatocytes per unit area were significantly ($p \le .0001$) reduced in Cr treatment as compared to control.



Fig. 2. Histological sections of mice testis in control (C), Cr treated (Cr), Morus (CrM), Jamun (CrJ) and combine co-postreatment of Morus and Jamun (CrMJ) groups

Abbas et al.; Eur. J. Nutr. Food. Saf., vol. 16, no. 1, pp. 66-78, 2024; Article no.EJNFS.111129



Fig. 3. Testicular smear with sperm morphology in control, Cr-treated and fruit extracts groups *C; control, Cr; chromium, t.s; testicular smear, Mr; Morus, J; Jamun, ol; oil emulsion, s.v; seminal vesicle fluid*

Table.	2. Sperm	micrometry.	ameliorative	effects of	of fruit e	xtracts c	on Cr	induced	anomalies

Parameters	Groups					
	С	Cr	Cr-M	C-J	Cr-MJ	
PBH CSA(µ²) of head ***	[†] 13.91±0.64ª	7.86±0.65 ^b	15.39±0.74ª	12.89±1.25ª	15.78±1.59ª	
PBH tail length (µ)*	85.57±12.11ª	75.92±12.05ª	102.10±5.45 ^b	85.99±3.25ª	86.84±1.70ª	
PBH Middle piece.	0.08±0.06ª	0.58±0.09 ^b	0.75±0.05°	0.95±0.14ª	1.05±0.05 ^d	
DM (µ)**						
CH CSA(µ ²) of	11.37±0.98ª	9.16±1.85ª	14.05±4.05ª	15.7±7.04ª	11.19±0.73ª	
head*						
CH tail length(µ)*	81.07±5.55ª	56.27±14.43 ^b	86.28±16.89 ^a	87.46±5.55ª	67.51±10.90 ^{ab}	
CH Middle piece.	0.96±0.12 ^a	0.39±0.15°	0.78±0.61 ^{ac}	0.67±0.08 ^{bc}	1.25±0.35 ^d	
DM (µ)*						

C: control. Cr: chromium treated, Cr-M: chromium+MFE, Cr-J: chromium+JFE, Cr-MJ: chromium+MFE+JFE. CSA: mean cross- sectional area of (PBH) sperm, DM: diameter (thickness) of parrot beak headed sperm and club headed (CH) sperm mid piece at 400X. *: p ≤ 0.05-0.01, **: p ≤ 0.001 ***: p ≤ .0001 {Statistical analysis (ANOVA: two factors without replication)}, [†] group means ±SEM, μ = μm, ^{a b c}: Anyone two groups not sharing a lower case letters differ significantly from each other (Duncan's Multiple Range comparison- post hoc analysis). n=10

4. DISCUSSION

Cr as environmental toxicant produce ROS and induce andro-hepatic anomalies evident by presence of necrosis and steatosis in testes and liver due to β -oxidation which divert metabolic pathway towards lipogenesis [53]. That process effect hepatocytes and kuffer cells to induce cirrhosis, dehydration and fibrosis. The lipid peroxidation interrupt steroid level and elevate testosterone. The ketonic bodies from fatty acids also drop pH which induce atrophy and biochemical changes (Figs. 2, 4) as Cd induce atrophy, testicular architecture disorganization and germinal epithelium disruption [54]. These process increases the acidity which ruptured the endothelial lining of testes indicate the sloughing and degeneration by the exposure of Cr as debris and dislodge spermatid in the lumen of ST, which altered their diameter which exactly co-related with [55] study about Cd toxicity. The primary hypertrophy with Cr exposures seems to cause accumulation of undifferentiated cells due to the lack of meiosis and spermatogenesis to produce spermatogenic arrest [56]. The necrosis of sertoli cells leading to the dislodgment of dead spermatic cells in ST due to the deficiency of nutrition, evident by significant elevation of CSA of ST may cause testicular hypoplasia and destruction of Leydig's cell; may cause infertility [57]. ROS by Cr exposure damage mitochondrial membrane permeability with deficiencv /insufficiency of sugar metabolism which causes the depletion of germinal epithelium in the ST and destruction of ST basement membrane (Fig. 2). Sertoli cells are interconnected by tight for blood-testis junctions; barrier, which temporarily permits the passage of spermatogenic cells and the loss of interstitial cells affects the process of spermatogenesis and spermiogenesis. Cr exposure detach the sperm heads and significant elevation of tail-less spermatozoa specify androgen receptor deformities durina terminal spermatids differentiation (Fig. 3, Tab; 2). The rodent sperms have one or more apical regular symmetrical hooks [58]. After Cr exposure the spermatozoa show the prominent anomalies like sperm head with irregular and wavy intermediate appearance (Fig. 3). Micrometric analysis (Tab; 2) of spermatozoa specified the alterations in CSA of sperm head, thickness of mid-piece, and length of the tail, that can be extrapolated with Cr related DNA damage and loss of polymerization of the micro tubular array into microtubules of the sperm tail [59]. The anabolic androgenic steroids ameliorated by drugs and can be the pharmacological products of plants like β-sitosterol and α-Tocopherol anthocyanin,

(vitamin E). Phytochemicals are the most important lipid-soluble antioxidants normalize chain reactions of lipid peroxidation to scavenge the free radicals [60]. The phytochemicals are best natural sugar and cholesterol regulators as compared to synthetic drugs [61] in the same token synchronized effect of Morus-Jamun ameliorate and improve the histological and micrometric cellular organization of testis and liver.

Apart from hepato-lipogenicity, atrophy, dehvdration, fibrosis and hepatocytic necrosis reported in this study oxidative stress contributed by Cr exposure. The defense against free radicals is associated with activities of SGOT, SGPT and bio-chelation of heavy metals from body. The environmental chemical exposure in the induction and progression of various diseases is significant [62]. The hyperlipidemia may indicate anomalies of anterior pituitary, liverbiliary dysfunction, and lipoprotein lipase cofactor deficiency corticosteroids [63]. LDL cholesterol and excess lipid with rise in HDL and VLDL damage liver and cause hepatic steatosis and displace the nucleus towards periphery. Histological studies of liver at micrometric level indicate the alteration central vein CSA in Cr treated group, due to accumulation of debris by



Fig. 4. Histology of Liver

The cross sectional of liver indicate hepatocytes, their nuclei and central vein CSA, Mean width (μm) of Sinusoidal Spaces (S.S) at (400x), Mean number of hepatocytes (N. Hepat) in 46225μ² area (100x), Mean relative area occupied by hepatocytes (R.A.Hepat) in 46225μ² area (100x)

PARAMETERS	GROUPS					
	С	Cr	Cr-M	Cr-J	Cr-MJ	
CSA (µ²) Hc***	† 237.95±12.56 ^a	187.39±19.19 ^b	143.14±5.58°	171.95±8.59 ^b	228.76±10.78ª	
CSA (µ²) H.N***	32.59±1.29ª	43.98±1.17 ^b	29.18±1.25°	39.88±1.87 ^d	32.89±0.75ª	
$CSA(\mu^2)$	2849.14	4213.55	2090.25	2250.24	6748.31	
C.V ***	±255.09ª	±773.89 ^b	±329.59ª	±201.58ª	±1466.29 °	
Width(µm)S.S***	6.17±0.63ª	7.25±0.83 ^b	5.18±0.66°	7.09±0.67 ^b	5.05±0.26°	
N. Hc /area ***	112.06±6.07ª	80.64±3.47 ^b	56.75±5.49°	95.13±4.45 ^d	85.89±0.09 ^b	
R.A.Hc /area***	26504.77 ±1457.93 ^a	12995.14 ±555.12 ^b	7593.72 ±722.14°	16380.55 ±699.12 ^d	19639.12 ±762.14°	

Table 3. Micrometry of mice liver, ameliorative effects of fruit extracts on Cr induced anomalies

C: control. Cr: chromium treated, Cr-M: chromium+MFE, Cr-J: chromium+JFE, Cr-MJ: chromium+MFE+JFE. Mean cross sectional area (µ²) of hepatocytes (Hc) at (400x), Mean CSA (µ²) of hepatocytic nucleus (H.N) at (400x), Mean CSA (µ²) of central vein (C.V) at (40x), Mean width (µm) of Sinusoidal Spaces (S.S) at (400x), Mean number of hepatocytes(N. Hepat) in 46225µ² area (100x), Mean relative area occupied by hepatocytes (R.A.Hepat) in 46225µ² area (100x). n=10. Statistical analysis (ANOVA: two factors without replication).*: p ≤ 0.05-0.01; **: p ≤ 0.001; ***: p ≤ .0001, [†] group means ±SEM, ^{a b c}: Anyone two groups not sharing a lower case letters differ significantly from each other (Duncan's Multiple Range comparison- post hoc analysis)



Abbas et al.; Eur. J. Nutr. Food. Saf., vol. 16, no. 1, pp. 66-78, 2024; Article no.EJNFS.111129

Fig. 5. Comprehensive summary of Ameliorative role of phytochemicals against Cr induced Andro-hepatic anomalies in mice

necrosis. The significant reduction in the CSA of central veins (CV) and number of hepatocytes while the elevation of hepatocytes nuclei size and sinusoidal spaces (SS) clearly intimate liver necrosis [64, 74]. Such changes also specify hypothyroidism, excess glucocorticoid and cortisol due to heavy metals exposure [65]. MFE and JFE like citrus fruits flavonoids protect the membrane integrity during RBC hemolysis [66].

In humans the stomach acts as a site for anthocyanin absorption from food, cyanidin-3glucoside and cyanidin-3, 5-diglucoside, penetrate into human's blood through liver cells [67]. The anthocyanin-rich extract decreases the lipid peroxidation; to normalize cholesterol by up regulating the peroxisome proliferator-activated receptors and influence the testosterone metabolism to ameliorate hyperglycemia and reproductive anomalies [68].

The ameliorative effect of melatonin also affects the spermatogenic proliferation in ST and metalloid exposure like vitamin-E reduces the oxidative damage induced by Pb [69]. Protective

activities of plants (deferiprone) act as an aluminum chelator and nullify the Al-induced toxicity [26]. Chemical compounds isolated from Morus nigra like anthocyanin, betulinic acid, flavonoid, β-sitosterol and germanicol, and from Syzygium cumini [68] are oleanolic acid, isoquercetin, quercetin, myricetin, tannins, delphinidin. petunidin. malvidin-dialucosides. ellagic acid, gallotannin, ellagitannin friedelin. betulinic acid, kaempferol, gallic acid, myricetine, pinocarveol, cineole, gerany-l-acetone, α-cadinol, eucarvone, muurolol and α-myrtenal have medicinal abilities and can be used in drugs [70].

The heavy metals induced testicular and hepatic histoarchitecture grievances which can be attenuated and chelated by using specific plants extracts as supplements [71]. Traditional medicinal plants are used in amelioration of diseases because plants components like antioxidants [68] have ability to chelate the heavy metals and organs can be recovered (Fig. 5) after chelation. Antioxidant have significant role against Cr induced oxidative stress by efficient chelation and radical scavenging ability during apoptosis. Natural fruit extracts are without any side effect and never alter the normal cecal microbial composition and intestinal microbiota; essential for carnitine palmitoyl transferase-1 pathway during amelioration [68,72]. The traditional herbal medicines are preferred than modern synthetic drugs and chimeric antigen receptor T-cell therapy against cancer [73,74] along with phytochemicals supplementations will ensure amelioration without side effects in future should be encouraged and sponsored.

5. CONCLUSION

There is no literature available for the cotreatment of Morus and Jamun, but these findings open the door and given results authorized findings about amelioration against heavy metals. The modern allopathic medicines have limited therapeutic options due to their huge serious side effects while the local herbal drugs must be used as an alternative way to cure diseases. Heavy metals like chromium is frequently used in cultural sweets and rice coloring but the hexavalent form should be banned after scientific authentication while Morus (Toot) and Jamun (Jambul) bio-products should be sponsored for clinical recommendation. There is need of another study to find out Morus and Jamun phytochemicals composition for cheapest medicinal formulation.

ACKNOWLEDGEMENTS

The authors are thankful to University of Sargodha and HEC Pakistan for providing support for conduction of this research work.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFRENCES

- 1. Asmatullah, Qureshi SN, Shakoori AR. Hexavalent chromium-induced congenital abnormalities in chick embryos. J Appl Toxicol. 1998;18(3):167-71.
- Mahmood A, Syed SS, Amna J, Mudassar S. Consumption: multivariate analysis and risk evaluation studies, exposure assessment of essential and potentially toxic metals in wheat-based sweets for human. Molecules. 2023;28(21):7365.
- 3. Stohs SJ, Bagchi D, Hassoun E, Bagchi M. Oxidative mechanism in the toxicity of

chromium and cadmium ions. J Environ Pathol Toxicol Oncol. 2001;20(2):77-88.

- 4. J J, S SS, R A, R G. Bioremediation of heavy metals using biosurfactants. Int J Biotechnol Appl. 2009;1(2):50-4.
- 5. Molinero O, Márquez S. Use of nutritional supplements in sports: Risks, knowledge, and behavioural-related factors. Nutr Hosp. 2009;24(2):128-34. PMID 19593480.
- 6. Thompson J, Bannigan J. Cadmium: toxic effects on the reproductive system and the embryo. Reprod Toxicol. 2008;25(3):304-15.
- 7. Monageng E, Offor U, Takalani NB, Mohlala K, Opuwari CS. A review on the impact of oxidative stress and medicinal plants on Leydig cells. Antioxidants (Basel). 2023;12(8):1559.
- 8. Coss CC, Bauler M, Narayanan R, Miller DD, Dalton JT. Alanine Aminotransferase Regulation by Androgens in Non-hepatic Tissues. Pharm Res. 2012;29(4):1046-56.
- 9. Keren I, Wu Y, Inocencio J, Mulcahy LR, Lewis K. Killing by bactericidal antibiotics does not depend on reactive oxygen species. Science. 2013;339(6124):1213-16.
- Veigas JM, Narayan MS, Laxman PM, Neelwarne B. Chemical nature, stability and bioef ficacies of anthocyanins from fruit peel of Syzygium cumini Skeels. Food Chem. 2007;105(2):619-27.
- 11. Veigas JM, Shrivasthava R, Neelwarne B. Efficient amelioration of carbon tetrachloride induced toxicity in isolated rat hepatocytes by Syzygium cumini Skeels extract. Toxicol In Vitro. 2008;22(6):1440-46.
- Jude CI, Catherine CI. Hepatoprotective effect of an extract of the rhizomes of Sansevieria Baker against CCl4 induced liver injury. Asian J Res Chem. 2011;4(12):1854-60.
- Awasthi M. Relevance of alkaline phosphatase activity of immobilized green algae and cyanobacteria for heavy metal toxicity monitoring. J Mater Environ Sci. 2012;3(3):446-51.
- Ali RFM. Effect of pomposia (Syzygium cumini) fruit juice on the stability of fried sunflower oil. J Food Technol. 2010;8(2):30-8.
- Ha KT, Yoon SJ, Choi DY, Kim DW, Kim JK, Kim CH. Protective effect of Lycium chinense fruit on carbon tetrachlorideinduced hepatotoxicity. J Ethnopharmacol. 2005;96(3):529-35.

- 16. Benherlal PS, Arumughan C. Chemical composition and in vitro antioxidant studies on *Syzygium cumini* fruit. J Sci Food Agric. 2007;87(14):2560-69.
- 17. Akunna GG, Ogunmodede OS, Saalu CL, Ogunlade B. Ameliorative effect of Moringa oleifera leaf extracts on Cr-induced testicular toxicity in rat testes. World J Life Sci Res. 2012;2(20):2249-74.
- Geetha S, Ram MS, Mongia SS, Singh V, Ilavazhagan G, Sawhney RC. Evaluation of antioxidant activity of leaf extract of Seabuckthorn on Cr(VI) induced oxidative stress in albino rats. Ethnopharmacol. 2003;87(2-3):247-51.
- 19. Mittal M, Flora SJS. Vitamin E supplementation protects oxidative stress during arsenic and fluoride antagonism in male mice. Drug Chem Toxicol. 2007;30(3):263-81.
- Rendon-Ramirez A, Cerbon-Solorzano J, Maldonado-Vega M. Vitamin-E reduces the oxidative damage on δ-aminolevulinic dehydratase induced by lead intoxication in rat erythrocytes. Toxicol Vitro. 2007;21:1121-26.
- 21. Narasimhacharya VR, Amaravadi L, Vasant RA. Amelioration of fluoride induced oxidative stress by Mangifera indica L. fruit. Spatula. D.D. 2011;1(4):181-88.
- 22. Glińska S, Gabara B. The effects of the anthocyanin-rich extract from red cabbage leaves on *Allium cepa* L. root tip cell ultrastructure. Ecotoxicol Environ Saf. 2011;74(1):93-8.
- 23. Demirdag K, Bakcecioglu IH, Ozercan IH. Role of L-carnitine in the prevention of acute liver damage induced by carbon tetrachloride in rats. J Gast Hepatol. 2004;19:333-38.
- 24. Xiao-Feng J, Qian J, Lu YH. The role of hepatoprotective effect of a flavonoid-rich extract of Salvia R.Br. on CCl4 induced acute hepatic injury in mice. J Med Plants Res. 2011;5(9):1558-63.
- 25. Ramanathan K, Anusuyadevi M, Shila S, Panneerselvam C. Ascorbic acid and tocopherol as potent modulators of apoptosis on arsenic induced toxicity in rats. Toxicol Lett. 2005;156(2): 297-306.
- 26. Albina ML, Bellés M, Sanchez DJ, Domingo JL. Evaluation of the protective activities of deferiprone, an Al– chelator, on Al-induced developmental toxicity. Teratology. 2000;62(2):86-92.

- Hussain T. Gupta RK. Sweetv K. Eswaran 27. Viiavakumar Μ. Rao CV. В. Nephroprotective activitv of Solanum xanthocarpum fruit extract against and gentamicin-induced nephrotoxicity renal dysfunction in experimental rodents. Asian Pac J Trop Med. 2012;5(9):686-91.
- Batool F, Sabir SM, Rocha JBT, Shah AH, Saify ZS, Ahmed SD. Evaluation of antioxidant and free radical scavenging activities of fruit extract from Zanthoxylum alatum: commonly used spices in Pakistan. Pak J Bot. 2010;42(6):4299-311.
- 29. Swami S, B, Singh JN. Thakor, Patil, M. Food Nutr Sci. Jamun (*Syzygium cumini*): A review of Its Food and Medicinal Uses. 2012;3:1100-17.
- Okon J, Esenowo G, Etim G, Umoh N. Phytochemical screening and haemopoetic study of the Extract of Baphia nitida on albino rats. Int. J. Mod. Biol Med. 2013;3(2):60-8.
- 31. Rout S, Banerjee R. Free radical scavenging, anti-glycation and tyrosinase inhibition properties of a polysaccharide fraction isolated from Punica granatum. Biores Technol. 2007;98(16):3159-63.
- 32. Ozsahin AD, Gokce Z, Yilmaz O, Kirecci OA. The fruit extract of three strawberry cultivars prevents lipid peroxidation and protects the unsaturated fatty acids in the Fenton reagent environment. Int J Food Sci Nutr. 2012;63(3):353-57.
- 33. Chitra M, Nithyanandhi K. Radical scavenging activity of *Trianthema triquetra* in male albino rats intoxicated with CCl4. J Environ Biol. 2007;28(2):283-85. PMID 17915766.
- 34. Palanivel MG, Rajkapoor B, Kumar RS, Einstein W. Hepatoprotective and antioxidant effect of Pisonia against CCL4induced hepatic damage in rats. Sci Pharm. 2008;76:203-15.
- 35. Shetty PC, Vidyasagar MS. Treatment of mice with leaf extract of Jamun protects against the radiation-induced damage in the intestinal mucosa of mice exposed to different doses of γ-radiation. Pharmacologyonline. 2008;1:169-95.
- Ali SS, Kasoju N, Luthra A, Singh A, Sharanabasava H, Sahu A, et al. Indian medicinal herbs as sources of antioxidants. Food Res Int. 2008;41(1):1-15.
- El-Sayed AM, Ezzat SM, Salama M, Sleem M, AA. Hepatoprotective and cytotoxic activities of Delonix regia flower extracts. Pharmacol J. 2011;3(19):49-56.

- Qin C, Li Y, Niu W, Ding Y, Zhang R, Shang X. Analysis and characterisation of anthocyanins in Mulberry Fruit. Czech J Food Sci. 2010;28(2):117-26.
- Volpato GT, Calderon IMP, Sinzato S, Campos KE, Rudge MV, Damasceno DC. Effect of Morus nigra aqueous extract treatment on the maternal-fetal outcome, oxidative stress status and lipid profile of streptozotocin-induced diabetic rats. J Ethnopharmacol. 2011;138(3):691-6.
- 40. Hamdy SM. Effect of Morus on enzymatic activities in rat. J Appl Sci Res. 2012;8(1):10-6.
- Pereira Braga C, Momentti AC, Barbosa Peixoto F, de Fátima Ferreira Baptista R, dos Santos FA, Fava FH et al. Influence of treatment with quercetin on lipid parameters and oxidative stress of pregnant diabetic rats. Can J Physiol Pharmacol. 2013;91(2):171-77.
- 42. Jagetia GC, Baliga MS. Evaluation of the radioprotective effect of the leaf extract of *Syzygium cumini* in mice exposed to a lethal dose of gamma-irradiation. Nahrung. 2003;47(3):181-5.
- 43. Goyal PK, Verma P, Sharma P. Evaluation of anticancer and anti-oxidative potential of Syzygium against BaP induced gastric carcinogenesis. Asian Pac J Cancer Prev. 2010;11:753-8.
- 44. Das S, Sarma G. Study of the hepatoprotective activity of the ethanolic extract of the pulp of Eugenia jambolana (Jamun) in albino rats. J Clin Diagn Res. 2009;3:1466-74.
- 45. Ayyanar M, Subash-Babu P. Syzygium cumini, a review of its phytochemical constituents and traditional uses. Asian Pac J Trop Biomed. 2012;2(3):240-6.
- 46. Wen L, Wu D, Jiang Y, Prasad KN, Lin S, Jiang G, et al. Identification of flavonoids in litchi (*Litchi chinensis*) Leaf and evaluation of anticancer activities. J Funct Foods. 2014;6:555-63.
- Qasemzadeh MJ, Sharifi H, Hamedanian M, M. The effect of Viola odorata Flower syrup on the cough of children with asthma: A double-blind, randomized controlled trial. J Evid Based Complement Altern Med. 2015;20(4):287-91.
- Ghazanfarpour M, Sadeghi R, Roudsari RL, Khorsand I, Khadivzadeh T, Muoio B. Red clover for treatment of hot flashes and menopausal symptoms: A systematic review and meta-analysis. J Obstet Gynaecol. 2016;36(3):301-11.

- 49. Guo Y, Zhi F, Chen P, Zhao K, Xiang H, Mao Q et al. Green tea and the risk of prostate cancer: A systematic review and meta-analysis. Medicine. 2017;96(13): e6426.
- Amira, Akacha T, Rebai., Lazhar, Zourgui. Mohamed, Amri. J Can Res Ther. Preventive effect of ethanolic extract of cactus (*Opuntia ficus-indica*) cladodes on methotrexate-induced oxidative damage of the small intestine in Wistar rats. 2018;14(10):779-84.
- 51. Al-Dabbagh B, Elhaty IA, Elhaw M, Murali C, Al Mansoori A, Awad B et al. Antioxidant and anticancer activities of chamomile (*Matricaria recutita*). BMC Res Notes. 2019;12(1):3.
- Zhang H, Liu Y, Wang L, Li Z, Zhang H, Wu J et al. Differential effects of estrogen/androgen on the prevention of nonalcoholic fatty liver disease in the male rat. J Lipid Res. 2013;54(2):345-57.
- 53. Damek-Poprawa M, Sawicka-Kapusta K. Damage to the liver, kidney, and testis with reference to burden of heavy metals in yellow-necked mice from areas around steelworks and zinc smelters in Poland. Toxicology. 2003;186(1-2):1-10.
- Sharma S, Vyas V, Tamot S, Manhor S. Histological changes in the testis of airbreathing fish, Heteropnuestes fossilis (Bloch) following Cd-chloride exposure. JCBPS. 2013;3(2):1216-21.
- Fabricia DSP, Aparecida M, Diamante S, Dolder H. Testis response to low dose of cadmium in rat. Int J Exp Pathol. 2010;91(2):125-31.
- Boekelheide K, Fleming SL, Johnson KJ, Patel SR, Schoenfeld HA. Role of Sertoli cells in injury-associated testicular germ cell apoptosis. Proc Soc Exp Biol Med. 2000;225(2):105-15.
- 57. Giagulli VA, Vermeulen A. Leydig cell function in infertile men with idiopathic oligospermic infertility. J Clin Endocrinol Metab. 1988;66(1):62-7.
- 58. Firman RC, Cheam LY, Simmons LW. Sperm competition does not influence sperm hook morphology in selection lines of house mice. J Evol Biol. 2011;24(4): 856-62.
- 59. Vijaya BR, Sasikala P, Karthik A, Shivakumar K. Sodium selenite induced oxidative stress and histological alterations in testis of albino mice. Int J Adv Sci Tech Res. 2013;3:2.

- Nah WH, Koh IK, Ahn HS, Kim MJ, Kang H, Jun JH, et al. Effect of Spirulina maxima on spermatogenesis and steroidogenesis in streptozotocin-induced type I diabetic male rats. Food Chem. 2012;134(1):173-9.
- 61. Rahmani R, Mahmoodi M, Karimi M, Hoseini F, Heydari R. The effect of capparis spinosa fruit hydroalcoholic extract on blood sugar and lipids in diabetic and normal rats. Zahedan J Res Med Sci. 2013;15(10):10.
- Kaiser JP, Lipscomb JC, Wesselkamper SC. Putative mechanisms of environmental chemical–induced steatosis. Int J Toxicol. 2013;32:3.
- Mccarty MF. An elevation of triglycerides reflecting decreased triglyceride clearance may not be pathogenic relevance to highcarbohydrate diets. Med. Hyp. 2004;63(6):1065-73.
- 64. Muthukumaravel K, Rajaraman P. A study on the toxicity of chromium on the histology of gill and liver of freshwater fish Labeo rohita. J Pure Appl Zool. 2013;1(2):122-6.
- 65. Chaumont A, Nickmilder M, Dumont X, Lundh T, Skerfving S, Bernard A. Associations between proteins and heavy metals in urine at low environmental exposures: Evidence of reverse causality. Toxicol Lett. 2012;210(3):345-52.
- 66. Heroor S, Beknal AK, Mahurkar N. Immunomodulatory activity of methanolic extracts of fruits and bark of Ficus in mice and on human neutrophils. Ind J Pharmacol. 2013;45(2):130-5.
- 67. Milbury PE, Vita JA, Blumberg JB. Anthocyanins are bioavailable in humans

following an acute dose of cranberry juice. J Nutr. 2010;140(6):1099-104.

- Naz N, Khan MR, Shabbir MA, Faisal MN. Effect of iron-fortified jamun leather on the Asunra-induced anemia in Sprague Dawley rats. Front Nutr. 2023;10: 1195981.
- 69. Penna-Videau S. Bustos-Obregon, E. Cermeno-Vivas, J. R., Chirino, D. Int J Morphol. Malathion affects spermatogenic proliferation in mouse. 2012;30(4):1399-407.
- 70. Kaneria M, Baravalia Y, Chanda S. Determination of antibacterial and antioxidant potential of some medicinal plants from Saurashtra Region. Ind J Pharm Sci. 2009;71:406-12.
- 71. Singh P, Deora K, Sankhla V, Mogra P. Curcumin rendered protection against cadmium chloride induced testicular damage in Swiss Albino mice. J Cell Mol Biol. 2012;10(2):31-8.
- 72. Koeth RA, Wang Z, Levison BS, Buffa JA, Org E, Sheehy BT et al. Intestinal microbiota metabolism of I-carnitine, a nutrient in red meat, promotes atherosclerosis. Nat Med. 2013;19(5):576-85.
- 73. Van SS, Kerre T. Chimeric antigen receptor T-cell therapy: Design improvements and therapeutic strategies in cancer treatment. Acta Clin Belg. 2018;13:1-7.
- 74. Khawja R, MZ, Hafiz M, Kausar R, Muhammad A, Tahir A. Effects of diazinon on the ovarian micro-anatomical and micrometric parameters of pregnant mice. Affarican J Biotechnol. 2011;10(65):14656-68.

© 2024 Abbas et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/111129