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Benefits of Carotenoid Astaxanthin: A Review

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Authors' contributions

This work was carried out in collaboration among all authors. Authors SD and SC planned and conceptualized the design of the article and wrote the first draft of the manuscript. Authors SR, AI and AB contributed substantially for the literature searches, correction and final article format. Authors SN, PM and FHR critically reviewed the manuscript and final refinement was done. All authors read and approved the final manuscript.

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Review Article

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ABSTRACT

Astaxanthin is a high value keto-carotenoid pigment renowned for its commercial application in various industries such as aquaculture, food, cosmetic, nutraceutical and pharmaceutical sectors. It is commonly employed in salmonid and crustacean aquaculture to give the pink hue that these species are known for. Scientific literature reviews have persistently demonstrated the instrumental role of astaxanthin in targeting several animal health conditions. Most importantly, the profound effect on pigmentation, where astaxanthin is frequently utilized as an additive in formulating diets to boost and improve the coloration of many aquaculture farmed species, subsequently product quality, consumers' acceptance and market demand are increasing, and revenue generated. Moreover, the wide range of other physiological benefits of astaxanthin includes various improvements in survival, growth performance, reproductive capacity, stress tolerance, and disease resistance as well. Also, astaxanthin has some other applications like, it is an anticancerous agent, it can prevent diabetes and cardiovascular diseases and enhances nutritional qualities. Astaxanthin products are used for commercial applications in the dosage forms as tablets, capsules, syrups, oils, soft gels, creams, biomass and granulated powders. Astaxanthin

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patent applications are available in food, feed and nutraceutical applications. This manuscript basically reviews the current available evidence regarding biological sources of astaxanthin, extraction procedures, stability, biological activity, health benefits, and commercial uses.

Keywords: Astaxanthin; extraction; health benefits; shrimp shell; biochemistry.

1. INTRODUCTION

Astaxanthin is a xanthophyll carotenoid, a red fat-soluble pigment. Some research showed that astaxanthin had better bio activity than other carotenoid pigments [1]. One of the major drug administrative body USFDA (The United States Food and Drug Administration) has approved the application of carotenoid pigment astaxanthin in animal and fish feed as a source of colouring agent [2-4]. Also, natural astaxanthin is recognized by The European Commission as a food dye. Since astaxanthin (3,3'-dihydroxy-4,4'diketo-,'-carotene) occurs naturally in large amounts and was initially discovered in the marine environment, it is mostly found in the flesh of salmonids, the carapace of crustaceans, and other marine organisms [5,6]. The naturally occurring carotenoid pigment astaxanthin is generally biosynthesized in microalgae at the primary production level of the food chain. Following the consumption of microalgae by crustaceans, zooplankton, or insects, astaxanthin is bioaccumulated to higher trophic levels where it is then swallowed by fish and other aquatic organisms. Astaxanthin is derived from H. pluvialis or acquired from seafood for use as a nutritional supplement in case of humans and other animals also. Astaxanthin ingestion can either prevent or lower the risk of several diseases in both humans and animals. This carotenoid pigment is best known as an essential aquacultural feed additive for imparting the pinkish-red coloration to the flesh of salmon, trout, ornamental fish, shrimp, lobsters and cravfish resulting in a better guality and acceptance of the consumer. Astaxanthin is increasingly being used as nutritional а

supplement in meals, feeds, nutraceuticals, and medications. Thus, the manuscript presents a thorough, in-depth, and current analysis of the literature that sheds light on the astaxanthin source, extraction processes, storage stability, biological activities, medical benefits in the treatment and prevention of various diseases, and commercial uses for mankind.

2. STRUCTURE OF ASTAXANTHIN

Astaxanthin is a pigment containing carbon, hydrogen and oxygen atoms $(C_{40}H_{52}O_4)$ with a molar mass of 596.84 g/mol (Fig. 1). Two terminal rings connected by a polyene chain make up astaxanthin. This molecule has two asymmetric carbons located at the 3. 3' positions of the β -ionone ring with hydroxyl group (-OH) on either end of the molecule. While one hydroxyl group reacts with a fatty acid, it forms monoester; whereas when both hydroxyl groups react with fatty acids it results di-ester. Astaxanthin, available natural sources, exists in in stereoisomers, geometric isomers, free and esterified forms among which the most prevalent stereoisomers in nature are (3S, 3'S) and (3R, Xanthophyllomyces 3'R). While veast dendrorhous creates the (3R, 3'R) isomer, Haematococcus synthesises the (3S, 3'S) isomer There are three isomers of synthetic [8]. astaxanthin are present: (3S, 3'S), (3R, 3'S), and (3R, 3'R). While the predominant astaxanthin stereoisomer in wild Atlantic salmon is (3S, 3'S), the Antarctic krill (Euphausia superba) contains (3R, 3'R), which is the principal stereoisomer primarily contains esterified form. [9]. Astaxanthin content (µg/g) in Antarctic krill, Copepod, Red yeast and Crab shell is shown in Fig. 2.

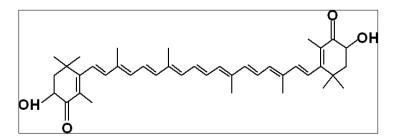


Fig. 1. Planner structure of astaxanthin [7]

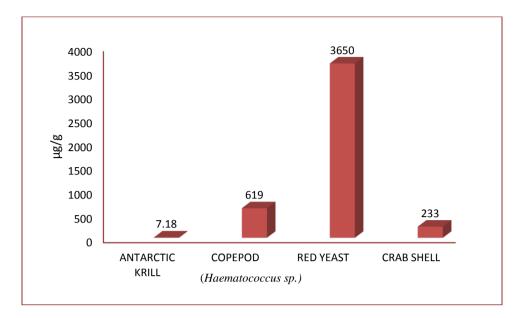


Fig. 2. Astaxanthin content (μ g/g) in Antarctic krill, Copepod, Red yeast and Crab shell [10,11,12,13]

3. SOURCES OF ASTAXANTHIN

The primary natural sources of astaxanthin are relatively simple microorganisms such as copepods, crab shell and yeast (Fig. 2). By ingesting astaxanthin containing creatures, animals do assemble astaxanthin in their tissues for beautiful colour, despite not being able to biochemically produce it. Salmonids and other fish, as well as organisms with exoskeletons like crabs, crayfish, lobsters, krill, and shrimp, consume zooplankton, which is fed on astaxanthin-rich algae in marine habitats; thus, astaxanthin is bioaccumulated and biomagnified subsequently at the higher trophic level (Table 2 & Fig. 3).

Class	Species	Astaxanthin (%) On a dry wt. basis	References
1. Chlorophyceae	Chlorococcum sp.	0.2 - 0.57	[14]
	C. zofingiensis	0.68 – 0.71	[15]
	H. pluvialis	4 - 7.72	[16]
2. Florideophyceae	Catenella repens	0.02	[17]
3. Alphaproteobacteria	Argobacterium	0.01	[18,19]
	auranticum Paracoccus sp.	2.2	
4. Tremellomycetes	Xanthophyllomyces dendrohous	0.41 – 0.97	[20, 21]

Table 1. Microbial sources of Astaxanthin

Table 2. Astaxanthin content of salmonids [22]

Fish Species	Astaxanthin (mg/kg flesh)
1. Atlantic salmon (Salmo salar)	3-10
2.Chinook salmon (Oncorhynchus tshawytscha)	5.4
3. Chum salmon (Oncorhynchus keta)	3-5
4. Coho salmon (Oncorhynchus kisutch)	10-21
5. Masu salmon (Oncorhynchus masou)	4.6
6. Pink salmon (Oncorhynchus gorbuscha)	4-7
7. Rainbow trout (Oncorhynchus mykiss)	24
8. Sockeye salmon (Oncorhynchus nerka)	26-38

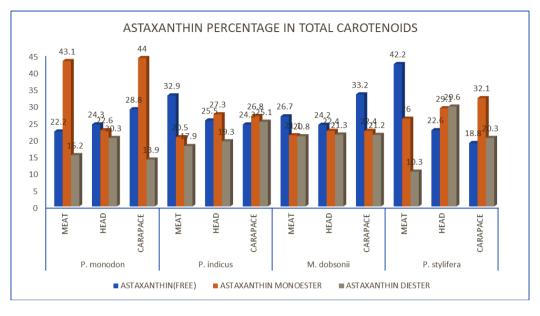


Fig. 3. Astaxanthin content (%) of shrimps [23]

4. EXTRACTION OF ASTAXANTHIN

4.1 Extraction of astaxanthin from *H. pluvialis*

climatic Under unfavorable conditions. astaxanthin accumulates mostly in the encysted cells of H. pluvialis up to 3-4% of the dry weight order to fully benefit from [24-26]. In astaxanthin's bioavailability, intact astaxanthinrich hematocysts must be mechanically disrupted before usage because of their thick and resistant cell walls [27-29]. Over the years, numerous methods have been created to disrupt H. pluvialis cells. Typically, physical or mechanical pretreatment-more specifically, bead milling and expeller pressing-is used to destroy cell walls [30-34]. A bead miller is a device that uses a disruption or milling chamber filled with tiny grinding beads (such as steel, glass, and ceramic) that are stirred up quickly and collide repeatedly. At these chambers, the dried biomass is fed, and compaction and shear forces

cause cell disruption in the bead impact zones. To prevent quality deterioration or spoilage, the algal biomass must be dehydrated as soon as possible using techniques like freeze-drying (lyophilization), spray-drying, drum-drying, and sun-drying [35-38]. Expeller pressing causes strong cell walls to burst by applying pressure and force simultaneously. The algal biomass must next be swiftly dehydrated by freeze-drying (lyophilization), spray-drying, drum-drying, and sun-drying to prevent quality degradation or spoilage [39].

A lipophilic pigment, astaxanthin can be digested in oils and organic solvents. Acids, organic solvents, and edible oils all have been used in a variety of ways to isolate astaxanthin from *H. pluvialis* (Table 3) [40,41]. Using several acid treatments at 70°C, Sarada et al. [40] assessed overall extraction efficiency of astaxanthin from *H. pluvialis* and reported that HCl treatment permitted 86-94 percent recovery of the pigment without changing its ester profile (Fig. 4).

Table 3. Extraction of astaxanthin by using acids, organic solvents and edible oils

Process Name	Procedure
1. Acid pre-treatment	According to Sarada et al. [40] 10 mg biomass was combined with 1 mL of HCl and incubated at 70°C for 2 minutes in a centrifugal tube. After the mixture had cooled, it was centrifuged for five minutes at 5000 rpm. It was then suspended in 1 mL of acetone after being rinsed twice in distilled water. The sample was collected after 20 minutes in the ice-water bath, and it was centrifuged by ultrasonography for 6 minutes at 4 degrees Celsius at 3500 rpm. In order to determine the HPLC of the extracted astaxanthin, the supernatants are then gathered. Nitrogen is used during the entire process, and light is avoided.

Process Name	Procedure
2. Solvents extraction	Binary organic solvents of hexane/isopropanol (6: 4) and 10 mg of the lyophilized organisms were added. After 20 minutes in an ice bath and an ultrasound-assisted auxiliary extraction, the sample was collected, centrifuged at 3500 rpm for 5 minutes at 4 C, then concentrated under vacuum [40]. On a dry basis, the collected yield is computed. In order to determine the HPLC of the extracted astaxanthin, the supernatants are then gathered. Nitrogen is used during the entire process, and light is avoided.
3. Methanol extraction	Sarada et al. [40] reported a 15 ml screw-top dark glass vial containing 10 mg of biomass was continuously sonicated in an ice bath with 1 ml each of methanol and acetone for 5 minutes. Samples were first extracted using 1 ml of methanol in a 15 ml dark glass vial with a screw top, and then centrifuged at 3500 rpm for 5 minutes at 4 °C. T hen it was used for HPLC analysis of the extracted astaxanthin. 1 ml of acetone was then added to a glass vial in this step. Nitrogen is used during the entire process, and light is avoided.
4. Oil-Soy Extraction	Sachindra and Mahendrakar [41] reported mix 2.5 g of cellular biomass with 20 ml of vegetable oil in a 250 ml flask (protected from light) and place on a hot plate with stirring at room temperature for 2 h. Further, it was filtered through cellulose (0.22 µm) to obtain an oil extract. Extractable astaxanthin using the supernatant was measured by HPLC.

4.2 Extraction of Astaxanthin from Shrimp Shell Waste

Table 4. Extraction of astaxanthin from shrimp	shell waste by using different solvents [42]
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Solvents	Procedure
1. Using 90% acetone	10 ml of 90% acetone was used to break 1 g of moist shrimp shell waste. Whatman filter paper was used to filter the extract. A colourless filtrate was obtained after the sample was once again proceeded with the same process using new solvent (3 times). 9.4 ml of 0.73 percent NaCl was added to the pooled extracts in a separate conical flask, and everything was thoroughly mixed. The epiphase was then thoroughly mixed and collected. The lower phase was mixed thoroughly with an equivalent volume of water before the epiphase was extracted.
2. Using Hexane, Isopropanol	Use 10 ml of hexane: isopropanol to pulverize 1 g of moist shrimp shell at a ratio of 3:2. It was done using Whatman No. 42 filter paper to prepare the extract. Until a colorless filtrate was achieved, samples were repeatedly extracted and filtered with fresh solvent. Hexane: isopropanol (3:2 v/v) mixed extracts were separated using an equal amount of a 1 percent (w/v) NaCl solution. The epi phase was obtained by collecting it and anhydrizing it over anhydrous sodium sulphate. The residue was then evaporated to dryness in vacuo and dissolved in 5 ml of hexane.
3. Using different Vegetable Oils (Coconut Oil, Palm Oil, Sunflower Oil)	1 g of moist shrimp shell was mixed with 10 ml of vegetable oil until a colorless sample was obtained. The ratio of oil: waste used for vegetable oil extraction was 2:1 for wet samples and 4:1 for dry samples. The solvent was removed in vacuo and redissolved in 5 ml of hexane. The antioxidant butylhydroxytoluene (BHT) was added at 0.05% (w/v) and heated at 70° C. for 150 minutes.

5. BIOCHEMISTRY OF ASTAXANTHIN

Conjugated double bonds, hydroxyl group and keto groups all are present in astaxanthin. It possesses hydrophilic and lipophilic characteristics [45]. The red hue is a result of the compound's conjugated double bonds, that operate as a potent antioxidant [46]. Due to its ability to bind cell membranes from the inside to the outside, astaxanthin demonstrated more biological activity than other antioxidants [47].

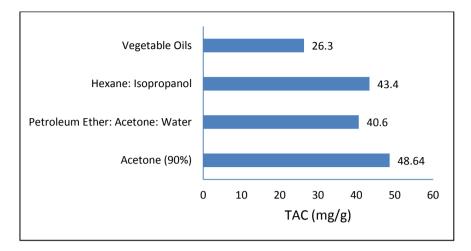


Fig. 4. Effect of the different extraction methods by using different solvents on total astaxanthin content (TAC) of extracts from shrimp shell waste [41,43,44]

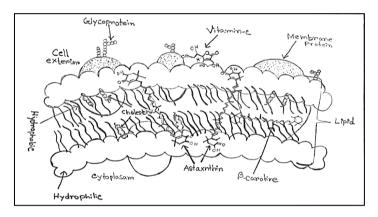


Fig. 5. Position of astaxanthin in the cell membrane

6. USES AND BENEFITS OF ASTAXANTHIN IN AQUATIC ANIMALS

6.1 Use in Reproduction

Astaxanthin is crucial towards the breeding as well as the cultivation of different aquaculture species. There are numerous proofs that indicate astaxanthin having a major effect on aquatic species' reproductive health, egg production, and egg quality [48-53]. In several fish farming facilities, efforts have always been made to enhance the quality of the eggs and larvae while nealecting the guality of the sperm. Therefore, it is imperative to comprehend the nutritional needs of farm animals since food availability directly affects a number of reproductive physiologic processes. Previous researches have shown supplementation astaxanthin how dietary affects the performance of diverse aquatic species' reproductive systems and brood stocks [51,52].

6.2 Use in Growth Parameters

Depending on production volume and cultivation techniques, feed often accounts for more than 60% of the overall hatchery management costs in aquaculture operations. In order to reduce production costs, it is crucial to produce feeds with nutrient components that support the growth and survival of the farmed species. The growth and survival (or both) of farmed fish and crustaceans during aquaculture techniques were positively correlated with dietary astaxanthin supplementation, according to a growing number of quantitative research publications [54,55].

6.3 Prevent Diseases

The rise of contagious diseases in intensive farming, particularly in the early phases of production, constitutes a substantial drawback or leading threat that has a considerable influence on the global economy. Farm animals are routinely subjected to physical stressors in highdensitv aquaculture operations. including grading. shipping, handling, vaccination. crowding and confinement, or any other physical disturbance that could be very stressful and immune-depressive. These unfavorable elements could disrupt the delicate equilibrium that aquatic creatures and their habitats must maintain, leading to stress reactions [56]. Excessive stress contributes to physiological malfunction, slower growth, immunosuppression, heightened susceptibility to pathogenic incursions, and even death. Therefore, it is crucial in aquaculture research to lessen unfavorable conditions that can cause a lot of stress and damage the host organism. Although farmed prawns and shrimp have been shown to exhibit extraordinary stress tolerance and disease resistance when compared to growth performance and survival, significant research efforts have been focused on reducing stress and improving immunity in aquaculture crustaceans and fish utilizing astaxanthin in their diet.

6.4 Use for Pigmentation

Aquaculture feed additives to improve the typical pinkish-red skin or meat coloration of aquatic species are perhaps astaxanthin's biggest potential use. The astaxanthin that is routinely added to their artificial diets and absorbed and deposited in rather high amounts causes skin and muscle coloration. From a commercial standpoint, maintaining natural pigmentation is crucial since it is closely linked to how customers perceive and interpret products, making it a crucial quality criterion before actual consumption that, in turn, drives up demand and product prices. The steadily growing aguaculture sector has created an insatiable demand for the carotenoid pigment since colour strongly influences customer preference and market demand for farmed species. Koi carp (Cyprinus carpio), kissing gourami (Helostoma temminckii), and other aquatic species have been the subject of studies examining the impact of dietary astaxanthin on the skin and flesh coloration of aquatic animals. [57,58,59].

7. USE OF ASTAXANTHIN IN FEED FORMULATION

The steadily growing aquaculture sector has created an insatiable demand for the carotenoid pigment since colour strongly influences customer preference and market demand for farmed species. Koi carp (*Cyprinus carpio*), kissing gourami (*Helostoma temminckii*), and other aquatic species have been the subject of studies examining the impact of dietary astaxanthin on the skin and flesh coloration of aquatic animals [60-63].

The single most crucial factor in the efficient use of intracellular astaxanthin during milling appears to be the disintegration or disruption of microalgal cells [33,64]. As a result, it has little effect on astaxanthin's stability [65]. However, grinding machinery, residence duration, and heat production all have a significant role in astaxanthin degradation. In order to ensure a consistent nutrient content in each fish pellet as part of the formulation, feed mixing is essential. However, mixing could introduce air into the mixture, which would lead to unfavorable carotene oxidation. To cope with air exposure and prevent air from entering the mixture, use a vacuum mixer. Alternately, it has been shown that the addition of secondary antioxidants (BHT and BHA) is effective in enhancing the oxidative stability of dietary carotenoids during feed processing [66].

Extrusion aims to reduce nutritional breakdown in food while increasing the digestibility of starch and protein [67-70]. Additionally, it is verified that the formulation is adjusted to generate floating or sinking pellets. While retention values in extruded feed ranged from 86% to 94%, astaxanthin was relatively constant across extrusion, with an average retention of 86% [65]. However, the stability of carotenoid pigments is most likely to be impacted by extrusion technology, which uses high amounts of heat, moisture, pressure, and mechanical shear. According to Storebakken et al. [71], the composition of astaxanthin during the manufacturing of extruded fish feed with a recovery range of 90-99 percent was not significantly impacted by the extruder temperatures of 102, 121, and 137°C. By improving the efficiency of nutrient consumption, pelletizing is the most popular process for formulating pellets [72,73]. To prevent the loss of pigment, the excess moisture is evaporated using a vacuum drying method (60-80°C temperature; a shelf-stable residual moisture of 10%). Additionally, a post-liquid coating application of fat or oil may lessen the chance of cooling immediately after potentially harming heat-sensitive carotenoids [74].

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8. STABILITY OF ASTAXANTHIN

Due to astaxanthin's inherent chemical instability. it's bioavailability has faced significant difficulties. which has limited its use as a functional food ingredient. This compels the market to think about fresh ideas for improving storage, stability, economic and efficient utilization of astaxanthin. Review of literatures reveals that, astaxanthin was stable at $70^{\circ}-90^{\circ}C$ in rice bran, gingelly (sesame), and palm oil with a retention of 84-90% of retention of astaxanthin content which can be used in food, pharmaceutical and nutraceutical applications, whereas astaxanthin content was reduced at 120 and 150 °C (Ranga). Anarjan and Tan [60] reported that degradation of astaxanthin was significantly higher in skimmed milk than orange juice. After 63 days of storage, only 10% of the astaxanthin dried at 180/110°C and kept at -21°C in nitrogen had degraded. At 4°C and 25°C, astaxanthin's storage stability was improved in a complex solution of hydroxypropyl-cyclodextrin and water. The optimal storage conditions for dry astaxanthin, according to Gouveia and Empis vacuum and [72]. were under nitrogen atmosphere in the dark, with high retention levels of more than 90% even after 18 months of storage. Astaxanthin stability was investigated polymeric microencapsulation with usina nanospheres, emulsions and β-cyclodextrin [75-77]. Encapsulating homogenised astaxanthinenriched H. pluvialis cells in a stiff polymer network of chitosan increased their durability when stored at -18°C in a nitrogen environment for 24 weeks with only an 8% of pigment degradation. [78].

9. EFFECTS OF ASTAXANTHIN ON HUMAN HEALTH

9.1 Antioxidant Property

Oxidation can be suppressed by antioxidants. Reactive oxygen species and free radicals start oxidative damage (ROS). These highly reactive chemicals are created by an organism's typical aerobic metabolism. Through chain reactions, excessively oxidative molecules can interact with proteins, lipids, and DNA, causing DNA damage and a variety of diseases [1]. Endogenous and exogenous antioxidants, like carotenoids with double bonds and polyene chains, can prevent this oxidative destruction by absorbing singlet oxygen and scavenging radicals to stop chain reactions. Carotenoids' biological advantages may derive from their antioxidant characteristics, which are linked to their chemical and physical associations with cell membranes. When compared to different carotenoids like lutein, lycopene, alpha- and beta-carotene, astaxanthin exhibited more antioxidant activity, according to research [79].

9.2 Prevents Lipid Peroxidation

Based on its unique chemical structure, astaxanthin can remain both inside and outside of the cell membrane. Compared to β -carotene and Vitamin C, which can be found inside the lipid bilayer, it offers superior protection [1]. It protects against oxidative damage through a number of ways, including quenching singlet oxygen, scavenging radicals to stop chain reactions, preserving membrane structure by preventing lipid peroxidation, boosting immune system performance, and controlling gene expression. Rats given ethanol to produce gastric ulcers and skin cancer displayed 80% anti-lipid peroxidation action with astaxanthin and its esters [1,80,81].

9.3 Anti-Diabetic Activity

People with diabetes mellitus typically have very high levels of oxidative stress. Due to the malfunctioning of pancreatic beta-cells and tissue damage in patients, it is brought on by hyperglycemia. Astaxanthin may reduce oxidative stress induced by hyperglycemia in pancreatic β -cells while simultaneously improving glucose and serum insulin levels [82].

9.4 Prevent Heart Diseases

Strong anti-oxidant astaxanthin has antiinflammatory effects in both humans and animals. Astaxanthin functions as a powerful therapeutic agent against the pathophysiological characteristics of atherosclerotic cardiovascular disease known as oxidative stress and inflammation [83].

9.5 Anticancerous Agent

The precise dose of antioxidants may be useful for rapid recognition of many degenerative disorders During regular aerobic metabolism, superoxide, hydrogen peroxide, and hydroxyl radical are produced. Singlet oxygen is formed by photochemical processes, whereas lipid peroxidation produces peroxyl radicals. Through the oxidation of DNA, proteins, and lipids, these oxidizing agents contribute to aging and degenerative disorders including cancer and atherosclerosis [84].

9.6 Effect on Immune System

Cells of the immune system are extremely vulnerable to damage by free radicals. Poly unsaturated fatty acids are found in the cell membrane. Being an antioxidant, astaxanthin resistance against provides free radical deterioration to maintain immune system defenses. There are studies on astaxanthin's impact on immunity in lab animals, but there aren't any for humans. In a mouse model, astaxanthin had more immunomodulatory effects than β-carotene [85].

9.7 Health Benefits

According to literature reviews, astaxanthin may have benefits for your health by preventing and conditions like cancer. chronic treating inflammatory diseases, metabolic syndrome, diabetes, diabetic nephropathy, cardiovascular gastrointestinal disorders, diseases. liver disorders, neurodegenerative disorders, eye skin disorders, exercise-induced disorders. fatique, and male infertility [86]. Currently, astaxanthin may be referred to as "a medical food" [1]. Although there are publications on how astaxanthin affects immunity in lab animals, there is a significant research deficit in the area of human clinical studies, and those publications are also few. As a result, there is a vast area for future study in the field of medicine related to the practical application, mechanism of action, and effectiveness of astaxanthin derived from Haematococcus species.

10. CONCLUSION

Currently, astaxanthin might be considered "a medicinal food." Although there are publications on how astaxanthin affects immunity in lab animals, there is a significant research deficit in the area of human clinical studies, and those publications are also few. As a result, there is a vast area for future study in the field of medicine about the use, mechanism, and effectiveness of astaxanthin. Astaxanthin ingestion can either prevent or lower the risk of a number of illnesses in both humans and animals. The pinkish-red colour of the flesh of salmon, trout, ornamental fish, shrimp, lobsters, and crayfish is imparted by this carotenoid pigment, which is well recognized as an important aquaculture feed addition. This results in greater quality and customer

acceptability. In foods, feeds, nutraceuticals, and medicines, astaxanthin used as a nutritional supplement has been expanding quickly. Future studies should concentrate on how astaxanthin esters affect various biological processes and how they are used in pharmaceutical and nutraceutical products.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Ranga Rao A, Siew Moi P, Sarada R, Ravishankar Gokare A. Astaxanthin: Sources, Extraction, Stability, Biological Activities and Its Commercial Applications. Mar Drugs. 2014;12:128-152. DOI: 10.3390/md12010128.
- Pashkow FJ, Watumull DG, Campbell CL. Astaxanthin: A novel potential treatment for oxidative stress and inflammation in cardiovascular disease. Am J Cardiol. 2008;101:58D–68D.

Available:https://doi.org/10.1016/j.amjcard. 2008.02.010

- 3. Zhuang X, Guo H, Alajlan N, Zhu H, Rabczuk T. Deep autoencoder based energy method for the bending, vibration, and buckling analysis of Kirchhoff plates with transfer learning. Eur J Mech-A/Solids. 2021;87:104225. Available:https://doi.org/10.1016/j.euromec hsol.2021.104225
- 4. Asker D, Awad T, Beppu T, Ueda K. A novel radio-toler- ant astaxanthinproducing bacterium reveals a new astaxanthin derivative: Astaxanthin dirhamnoside. Methods Mol Biol. 2012; 892:61–97.

DOI: 10.1007/978-1-61779-879-5_4

- 5. Han D, Li Y, Hu Q. Astaxanthin in microalgae: pathway, functions and biotechnological implications. Algae. 2013; 28:131–147.
- Begum H, Yusoff FM, Banerjee S, Khatoon H, Shariff M. Availability and utilization of pigments from microalgae. Crit Rev Food Sci Nutr. 2016;56:2209–2222. Available:https://doi.org/10.1080/10408398 .2013.764841.
- 7. Stachowiak B, Szulc P. Astaxanthin for the food industry. Molefw. 2021;(9):2666.

Available:https://doi.org/10.3390/molecules 26092666

8. Hussein G, Sankawa U, Goto H, Matsumoto K, Watanabe H. Astaxanthin, a carotenoid with potential in human health and nutrition. J Nat Prod. 2006;69(3):443-9.

> Available:https://doi.org/10.1021/np050354 +

 Foss P, Storebakken T, Austreng E, Liaaenjensen S. Carotenoids in diets for salmonids: V. Pigmentation of rainbow trout and sea trout with astaxanthin and astaxanthin dipalmitate in comparison with canthaxanthin. Aquac. 1987;65(3-4):293-305.

Available:https://doi.org/10.1016/0044-8486(87)90242-0

 Cong XY, Miao JK, Zhang HZ, Sun WH, Xing LH, Sun LR, Zu L et al. Effects of drying methods on the content, structural isomers, and composition of astaxanthin in antarctic krill. ACS omega. 2019;4(19): 17972-80.

Available:https://doi.org/10.1021/acsomeg a.9b01294

11. Caramujo MJ, De Carvalho CC, Silva SJ, Carman KR. Dietary carotenoids regulate astaxanthin content of copepods and modulate their susceptibility to UV light and copper toxicity. Mar Drugs. 2012;10(5): 998-1018.

https://doi.org/10.3390/md10050998.

- 12. Choi SK, Kim JH, Park YS, Kim YJ, Chang HI. An efficient method for the extraction of astaxanthin from the red yeast *Xanthophyllomyces dendrorhous*. J Microbiol Biotechnol. 2007;17(5):847-52.
- Rodrigues LA, Pereira CV, Leonardo IC, Fernández N, Gaspar FB, Silva JM et al. Terpene-Based natural deep eutectic systems as efficient solvents to recover astaxanthin from brown crab shell residues. ACS Sustain Chem Eng. 2020;8(5):2246-59. Available:https://doi.org/10.1021/acssusch emeng.9b06283.
- 14. Zhang DH, ML, Phang SM. Ng Composition and accumulation of secondary carotenoids in Chlorococcum J Appl Phycol. 1997;9:147-155. sp. Available:https://doi.org/10.1023/A:100792 6528388.
- 15. Orosa M, Valero JF, Herrero C, Abalde J. Comparison of the accumulation of

astaxanthin in *Haematococcus pluvialis* and other green microalgae under Nstarvation and high light conditions. Biotechnol Lett. 2001;13:1079-85. Available:https://doi.org/10.1023/A:101051 0508384.

- Lee YK, Ding SY. Cell cycle and accumulation of astaxanthin in *Haematococcus lacustris* (Chlorophyta) 1. J Phycol. 1994;(3):445-9. Available:https://doi.org/10.1111/j.0022-3646.1994.00445
- Banerjee K, Ghosh R, Homechaudhuri S, Mitra A. Bio- chemical composition of marine macroalgae from Gangetic delta at the apex of Bay of Bengal. Afr J Basic Appl Sci. 2009;1:96–104.
- Yokoyama A, Adachi K, Shizuri Y. New carotenoid gluco- sides, astaxanthin glucoside and adonixanthin glucoside, isolated from the astaxanthin producing marine bacterium, *Agrobacterium aurantiacum.* J Nat Prod. 1995;58:1929–1933

Available:https://doi.org/10.1021/np50126a 022

- EFSA. Safety and efficacy of panaferd-AX (red carote- noid-rich bacterium *Paracoccus carotinifaciens*) as feed additive for salmon and trout. EFSA J. 2007;546:1–30.
- Rodríguez-Sáiz M, de la Fuente JL, Barredo JL. Xanthophyllomyces dendrorhous for the industrial production of astaxanthin. Appl Microbiol Biotechnol. 2010;88(3):645-658. Available:https://doi.org/10.1007/s00253-

010-2814-x

- Gassel S, Schewe H, Schmidt I, Schrader J, Sandmann G. Multiple improvement of astaxanthin biosynthesis in *Xanthophyllomyces dendrorhous* by a combination of conventional mutagenesis and metabolic pathway engineering. Biotechnol Lett. 2013;(4):565-9. Available:https://doi.org/10.1007/s10529-012-1103-4
- 22. EFSA (European Food Safety Authority). Opinion of the scientific panel on additives and products or substances used in animal feed on the request from the European commission on the safety of use of colouring agents in animal human nutrition. EFSA J. 2005;291:1–40.

- Sachindra NM, Bhaskar N, Mahendrakar NS. Carotenoids in different body components of Indian shrimps. J Sci Food Agric. 2005;85(1):167-72. Available:https://doi.org/10.1002/jsfa.1977
- 24. Lemoine Y, Schoefs B. Secondary ketocarotenoid astaxanthin biosynthesis in algae: A multifunctional response to stress. Photosynth Res. 2010;06(1):155-77. Available:https://doi.org/10.1007/s11120-010-9583-3
- 25. Wayama M, Ota S, Matsuura H, Nango N, Hirata A, Kawano S. Three-dimensional ultrastructural study of oil and astaxanthin accumulation during encystment in the green alga *Haematococcus pluvialis*. PloS one. 2013;8(1):53618.

Available:https://doi.org/10.1371/journal.po ne.0053618

- Chekanov K, Lobakova E, Selyakh I, Semenova L, Sidorov R, Solovchenko A. Accumulation of astaxanthin by a new *Haematococcus pluvialis* strain BM1 from the White Sea coastal rocks (Russia). Mar. Drugs. 2014;12(8): 4504-20. Available:https://doi.org/10.3390/md12084 504
- Damiani E, Rosati L, Castagna R, Carloni P, Greci L. Changes in ultraviolet absorbance and hence in protective efficacy against lipid peroxidation of organic sunscreens after UVA irradiation. J Photochem Photobiol B. 2006;82(3):204-13.

Available:https://doi.org/10.1016/j.jphotobio I.2005.03.011

28. Kaczor A, Baranska M. Structural changes of carotenoid astaxanthin in a single algal cell monitored in situ by Raman spectroscopy. Anal Chem. 2011;83(20): 7763-70.

Available:https://doi.org/10.1021/ac201302 f

 Kim D, Vijayan D, Ramasamy P, Oh Y. Cell-wall disrup- tion and lipid/astaxanthin extraction from microalgae: Chlorella and Haematococcus. Bioresour Technol. 2016; 199:300–310. Available:https://doi.org/10.1016/j.biortech.

Available:https://doi.org/10.1016/j.biortech. 2015.08.107

 Mercer P, Armenta RE. Developments in oil extraction from microalgae. Eur J Lipid Sci Technol. 2011;113(5):539-47. Available:https://doi.org/10.1002/ejlt.20100 0455

 Razon LF, Tan RR. Net energy analysis of the production of biodiesel and biogas from the microalgae: *Haematococcus pluvialis* and Nannochloropsis. Appl Energy. 2011; 88(10):3507-14.

Available:https://doi.org/10.1016/j.apenerg y.2010.12.052

 Zhang J, Lü F, Shao L, He P. The use of biochar-amended composting to improve the humification and degradation of sewage sludge. Bioresour Technol. 2014; 168:252-8.

Available:https://doi.org/10.1016/j.biortech. 2014.02.080

 Cuellar-Bermudez SP, Aguilar-Hernandez I, Cardenas-Chavez DL, Ornelas-Soto N, Romero-Ogawa MA, Parra-Saldivar R. Extraction and purification of highvalue metabolites from microalgae: Essential lipids, astaxanthin and phycobiliproteins. Microb Biotechnol. 2015; (2):190-209.

Available:https://doi.org/10.1111/1751-7915.12167

- Kim Y, Rush AM. Sequence-level knowledge distillation. arXiv preprint. 2016; arXiv:1606.07947. Available:https://doi.org/10.48550/arXiv.16 06.07947
- Doucha J, Lívanský K. Influence of processing parameters on disintegration of Chlorella cells in various types of homogenizers. Appl. Microbiol. Biotechnol. 2008;(3):431-40.

Available:https://doi.org/10.1007/s00253-008-1660-6

 Schwenzfeier A, Wierenga PA, Gruppen H. Isolation and characterization of soluble protein from the green microalgae *Tetraselmis sp.* Bioresour. Technol. 2011;102(19):9121-9127. Available:https://doi.org/10.1016/j.biortech.

Available:https://doi.org/10.1016/j.biortech. 2011.07.046.

- Postma DS, Bush A, van den Berge M. Risk factors and early origins of chronic obstructive pulmonary disease. The Lancet. 2015;385(9971):899-909. Available:https://doi.org/10.1016/S0140-6736(14)60446-3.
- Günerken E, D'Hondt E, Eppink M, Elst K, Wijffels R. Influence of nitrogen depletion in the growth of N. oleoabundans on the

release of cellular components after bead milling. Bioresour Technol. 2016;214:89-95.

Available:https://doi.org/10.1016/j.biortech. 2016.04.072

 Shah MM, Liang Y, Cheng JJ, Daroch M. Astaxanthin-producing green microalga *Haematococcus pluvialis*: From single cell to high value commercial products. Front Plant Sci. 2016;7 531.

vhttps://doi.org/10.3389/fpls.2016.00531.

40. Sarada R, Vidhyavathi R, Usha D, Ravishankar GA. An efficient method for extraction of astaxanthin from green alga *Haematococcus pluvialis.* J Agric Food Chem. 2006;54(20):7585-8.

Available:https://doi.org/10.1021/jf060737t

- 41. Sachindra NM, Mahendrakar NS. Process optimization for extraction of carotenoids from shrimp waste with vegetable oils. Bioresour Technol. 2005;96(10)1195-200. Available:https://doi.org/10.1016/j.biortech. 2004.09.018
- 42. Sindhu R, Kuttiraja M, Binod P, Janu KU, Sukumaran RK, Pandey A. Dilute acid pretreatment and enzymatic saccharification of sugarcane tops for bioethanol production. Bioresour Technol. 2011;102(23):10915-21.

Available:https://doi.org/10.1016/j.biortech. 2011.09.066

 Kelley JM, Emerson SU, Wagner RR. The glycoprotein of vesicular stomatitis virus is the antigen that gives rise to and reacts with neutralizing antibody. J Virol. 1972;10(6):1231-5. Available:https://doi.org/10.1128/jvi.10.6.12

Available:https://doi.org/10.1128/jvi.10.6.12 31-1235.1972

44. Vimala K, Sivudu KS, Mohan YM, Sreedhar B, Raju KM. Controlled silver nanoparticles synthesis in semi-hydrogel networks of poly (acrylamide) and carbohydrates: A rational methodology for antibacterial application. Carbohydr Polym. 2009;75(3):463-71.

Available:https://doi.org/10.1016/j.carbpol. 2008.08.009.

45. Higuera-Ciapara I, Felix-Valenzuela L, Goycoolea FM. Astaxanthin: A review of its chemistry and applications. Crit Rev Food Sci Nutr. 2006;46(2):185-96.

Available:https://doi.org/10.1080/10408690 590957188

46. Guerin M, Huntley ME, Olaizola M. Haematococcus astaxanthin: Applications for human health and nutrition. Trends Biotechnol. 2003;21(5): 210-6.
Available:https://doi.org/10.1016/S0167-

Available:https://doi.org/10.1016/S0167-7799(03)00078-7.

- 47. Yuan Y, Zhao B, Zhou S, Zhong S, Zhuang L. Electrocatalytic activity of anodic biofilm responses to pH changes in microbial fuel cells. Bioresour Technol. 2011;102(13): 6887-91. https://doi.org/10.1016/j.biortech.2011.04.0 08
- Vassallo-Agius R, Imaizumi H, Watanabe T, Yamazaki T, Satoh S, Kiron V. The influence of astaxanthin supplemented dry pellets on spawning of striped jack. Fish Sci. 2001;67(2):260-70. Available:https://doi.org/10.1046/j.1444-2906.2001.00248.x
- Ahmadi MR, Bazyar AA, Safi S, Ytrestøyl T, Bjerkeng B. Effects of dietary astaxanthin supplementation on reproductive characteristics of rainbow trout (Oncorhynchus mykiss). J Appl Ichthyol. 2006;22(5):388-94. Available:https://doi.org/10.1111/j.1439-

0426.2006.00770.x

 Paibulkichakul C, Piyatiratitivorakul S, Sorgeloos P, Menasveta P. Improved maturation of pond-reared, black tiger shrimp (*Penaeus monodon*) using fish oil and astaxanthin feed supplements. Aquac. 2008;282(1-4):83-9. Available:https://doi.org/10.1016/j.aquacult

ure.2008.06.006 51. Et A, Tizkar B. The Effects of Dietary Supplementation of Astaxanthin and.

Caspian J Environ Sci. 2013;11(2):217-31.
52. Tizkar B, Kazemi R, Alipour A, Seidavi A, Naseralavi G, Ponce-Palafox JT. Effects of dietary supplementation with astaxanthin and β-carotene on the semen quality of real-field.

goldfish (*Carassius auratus*). Theriogenology. 2015;84(7): 1111-7.

53. Palma J, Andrade JP, Bureau DP. The impact of dietary supplementation with astaxanthin on egg quality and growth of long snout seahorse (*Hippocampus guttulatus*) juveniles. Aquac Nutr. 2017; 23(2):304-12.

Available:https://doi.org/10.1111/anu.1239

- Kumar V, Pillai BR, Sahoo PK, Mohanty J, Mohanty S. Effect of dietary astaxanthin on growth and immune response of Giant freshwater prawn *Macrobrachium rosenbergii* (de man). Asian Fish Sci. 2009;22(1):61-9.
- Niu J, Tian LX, Liu YJ, Yang HJ, Ye CX, Gao W, et al. Effect of dietary astaxanthin on growth, survival, and stress tolerance of post larval shrimp, Litopenaeus vannamei. J World Aquac Soc. 2009;40(6):795-802.

Available:https://doi.org/10.1111/j.1749-7345.2009.00300.x

56. Scholthof HB. Heterologous expression of viral RNA interference suppressors: RISC management. Plant Physiol. 2007;145(4): 1110-7.

Available:https://doi.org/10.1104/pp.107.10 6807.

57. Lim KC, Yusoff FM, Shariff M, Kamarudin MS. Astaxanthin as feed supplement in aquatic animals. Rev Aquac. 2018 Aug;10(3):738-73.

Available:https://doi.org/10.1111/raq.12200

- Kopecký J. The Effect of Astaxanthin and β-carotene on the Colour of the Kissing Gourami (*Helostoma temminckii*). Acta Fytotech Zootech. 2013;16(3).
- 59. Nguyen VP, Kim SW, Kim H, Kim H, Seok KH, Jung MJ et al. Biocompatible astaxanthin as a novel marine-oriented agent for dual chemo-photothermal therapy. PloS one. 2017;12(4):e0174687. Available:https://doi.org/10.1371/journal.po ne.0174687
- Anarjan N, Tan CP. Developing a three component stabilizer system for producing astaxanthin nanodispersions. Food Hydrocoll. 2013;30(1):437-47. Available:https://doi.org/10.1016/j.foodhyd. 2012.07.002
- 61. Bustos-Garza C, Yáñez-Fernández J, Barragán-Huerta BE. Thermal and pH stability of spray-dried encapsulated astaxanthin oleoresin from *Haematococcus pluvialis* using several encapsulation wall materials. Food Res Int. 2013;54(1):641-9.

Available:https://doi.org/10.1016/j.foodres. 2013.07.061.

62. De Bruijn WJ, Weesepoel Y, Vincken JP, Gruppen H. Fatty acids attached to alltrans-astaxanthin alter its cis-trans equilibrium, and consequently its stability, upon light-accelerated autoxidation. Food Chem. 2016;194:1108-15.

Available:https://doi.org/10.1016/j.foodche m.2015.08.077

- Martínez-Delgado AA, Khandual S, Villanueva–Rodríguez SJ. Chemical stability of astaxanthin integrated into a food matrix: Effects of food processing and methods for preservation. Food Chem. 2017;225: 23-30. Available:https://doi.org/10.1016/j.foodche
- m.2016.11.092
 64. Kim DY, Vijayan D, Praveenkumar R, Han JI, Lee K, Park JY et al. Cell-wall disruption and lipid/astaxanthin extraction from microalgae: Chlorella and Haematococcus. Bioresour. Technol. 2016;199:300-10. Available:https://doi.org/10.1016/j.biortech. 2015.08.107
- 65. Anderson JS, Sunderland R. Effect of extruder moisture and dryer processing temperature on vitamin C and E and astaxanthin stability. Aquac. 2002;207(1-2):137-49.

Available:https://doi.org/10.1016/S0044-8486(01)00787-6

- Jintasataporn O, Yuangsoi B. Stability of carotenoid diets during feed processing and under different storage conditions. Molefw. 2012;17(5):5651-60. Available:https://doi.org/10.3390/molecules 17055651.
- Glencross BD, Tocher DR, Matthew C, Gordon Bell J. Interactions between dietary docosahexaenoic acid and other longchain polyunsaturated fatty acids on performance and fatty acid retention in post-smolt Atlantic salmon (*Salmo salar*). Fish Physiol Biochem. 2014;40(4):1213-27.

Available:https://doi.org/10.1007/s10695-014-9917-8

- 68. Glencross BD, Bermudes M. Adapting bioenergetic factorial modelling to understand the implications of heat stress on barramundi (*Lates calcarifer*) growth, feed utilisation and optimal protein and energy requirements-potential strategies for dealing with climate change? Aquac Nutr. 2012;18(4):411-22. Available:https://doi.org/10.1111/j.1365-2095.2011.00913.x
- 69. Kamarudin MS, De Cruz CR, Saad CR, Romano N, Ramezani-Fard E. Effects of extruder die head temperature and pre-

gelatinized taro and broken rice flour level on physical properties of floating fish pellets. Anim Feed Sci Technol. 2018;236:122-30.

Available:https://doi.org/10.1016/j.anifeeds ci.2017.12.007

- 70. Salunkhe DK, Bolin HR, Reddy NR. Storage, processing, and nutritional quality of fruits and vegetables. Volume I. Fresh fruits veg. CRC press; 1991.
- 71. Storebakken T, Sørensen M, Bjerkeng B, Harris J, Monahan P, Hiu S. Stability of astaxanthin from red yeast, Xanthophyllomyces dendrorhous, during feed processing: effects of enzymatic cell wall disruption and extrusion temperature. Aquac. 2004;231(1-4):489-500.

Available:https://doi.org/10.1016/j.aquacult ure.2003.10.034

72. Gouveia L, Empis J. Relative stabilities of microalgal carotenoids in microalgal extracts, biomass and fish feed: effect of storage conditions. Innov Food Sci Emerg Technol. 2003;4(2):227-33.

Available:https://doi.org/10.1016/S1466-8564(03)00002-X

73. Jintasataporn O, Yuangsoi B. Stability of carotenoid diets during feed processing and under different storage conditions. Molefw. 2012;17(5):5651-60.

Available:https://doi.org/10.3390/molecules 17055651

- 74. Boon CS, McClements DJ, Weiss J, Decker EA. Factors influencing the chemical stability of carotenoids in foods. Crit Rev Food Sci Nutr. 2010;50(6):515-32. Available:https://doi.org/10.1080/10408390 802565889
- Higuera-Ciapara I, Felix-Valenzuela L, Goycoolea FM, Argüelles-Monal W. Microencapsulation of astaxanthin in a chitosan matrix. Carbohydr Polym. 2004; 56(1):41-5. Available:https://doi.org/10.1016/j.carbpol.

2003.11.012

- Tachaprutinun A, Udomsup T, Luadthong C, Wanichwecharungruang S. Preventing the thermal degradation of astaxanthin through nanoencapsulation. Int J Pharm. 2009;374(1-2):119-24. Available:https://doi.org/10.1016/j.ijpharm. 2009.03.001
- 77. Chen X, Chen R, Guo Z, Li C, Li P. The preparation and stability of the inclusion

complex of astaxanthin with β-cyclodextrin. Food Chem. 2007;101(4):1580-4. Available:https://doi.org/10.1016/j.foodche m.2006.04.020

- Kittikaiwan P, Powthongsook S, Pavasant P, Shotipruk A. Encapsulation of *Haematococcus pluvialis* using chitosan for astaxanthin stability enhancement. Carbohydr polym. 2007;70(4):378-85. Available:https://doi.org/10.1016/j.carbpol. 2007.04.021.
- Naguib YM. Antioxidant activities of astaxanthin and related carotenoids. J Agric Food Chem. 2000;48(4):1150-4. Available:https://doi.org/10.1021/jf991106k
- Rao AR, Baskaran V, Sarada R, Ravishankar GA. In vivo bioavailability and antioxidant activity of carotenoids from microalgal biomass—A repeated dose study. Food Res Int. 2013;54(1):711-7.

Available:https://doi.org/10.1016/j.foodres. 2013.07.067

 Kamath BS, Srikanta BM, Dharmesh SM, Sarada R, Ravishankar GA. Ulcer preventive and antioxidative properties of astaxanthin from *Haematococcus pluvialis*. European J Pharm Pharmacol. 2008; 590(1-3):387-95.
 Available:https://doi.org/10.1016/i.einbar.2

Available:https://doi.org/10.1016/j.ejphar.2 008.06.042

- 82. Uchiyama K, Naito Y, Hasegawa G, Nakamura N, Takahashi J, Yoshikawa T. Astaxanthin protects β-cells against glucose toxicity in diabetic db/db mice. Redox Report. 2002;7(5):290-3. Available:https://doi.org/10.1179/13510000 2125000811
- Fassett RG, Coombes JS. Astaxanthin: A potential therapeutic agent in cardiovascular disease. Mar Drugs. 2011;9(3):447-65.

Available:https://doi.org/10.3390/md90304 47.

 Ryu SK, King TJ, Fujioka K, Pattison J, Pashkow FJ, Tsimikas S. Effect of an oral astaxanthin prodrug (CDX-085) on lipoprotein levels and progression of atherosclerosis in LDLR-/- and ApoE-/mice. Atherosclerosis. 2012;222(1):99-105.

Available:https://doi.org/10.1016/j.atheroscl erosis.2012.02.002

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- Jyonouchi H, Sun S, Tomita Y, Gross MD. Astaxanthin, a carotenoid without vitamin A activity, augments antibody responses in cultures including T-helper cell clones and suboptimal doses of antigen. J Nutr. 1995;125(10):2483-92. Available:https://doi.org/10.1093/jn/125.10. 2483
- Yuan C, Jin Z, Xu X. Inclusion complex of astaxanthin with hydroxypropyl-βcyclodextrin: UV, FTIR, 1H NMR and molecular modeling studies. Carbohydr Polym. 2012;89(2):492-6. Available:https://doi.org/10.1016/j.carbpol. 2012.03.033

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